Extensions to the Cox Model Time Dependent Covariates

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Bone Marrow Transplant Data

- Copelan et al. (1991) study of allogenic bone marrow transplant therapy for acute myeloid leukemia (AML) and acute lymphoblastic leukemia (ALL).
- **Possible intermediate events are graft vs. host** disease (GVHD), an immunological rejection response to the transplant, and platelet recovery, a return of platelet count to normal levels. One or the other, both in either order, or neither may occur.
- \blacksquare End point events are relapse of the disease or death.
- Any or all of these events may be censored.

KMsurv bmt data

The bmt data frame has 137 rows and 22 columns.

This data frame contains the following columns:

- group Disease Group 1-ALL, 2-AML Low Risk, 3-AML High Risk
- t1 Time To Death Or On Study Time
- t2 Disease Free Survival Time (Time To Relapse, Death Or End Of Study)
- d1 Death Indicator 1-Dead 0-Alive
d2 Belanse Indicator 1-Belansed (
- Relapse Indicator 1-Relapsed, 0-Disease Free
- d3 Disease Free Survival Indicator 1-Dead Or Relapsed, 0-Alive Disease Free)
- ta Time To Acute Graft-Versus-Host Disease
- da Acute GVHD Indicator 1-Developed Acute GVHD 0-Never Developed Acute GVHD)
- tc Time To Chronic Graft-Versus-Host Disease
- dc Chronic GVHD Indicator 1-Developed Chronic GVHD

0-Never Developed Chronic GVHD

- tp Time To Platelet Recovery
- dp Platelet Recovery Indicator 1-Platelets Returned To Normal,
	- 0-Platelets Never Returned to Normal

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KMsurv bmt data

- z1 Patient Age In Years
- z2 Donor Age In Years
- z3 Patient Sex: 1-Male, 0-Female
- z4 Donor Sex: 1-Male, 0-Female
- z5 Patient CMV Status: 1-CMV Positive, 0-CMV Negative
- z6 Donor CMV Status: 1-CMV Positive, 0-CMV Negative
- z7 Waiting Time to Transplant In Days
- z8 FAB: 1-FAB Grade 4 Or 5 and AML, 0-Otherwise
- z9 Hospital: 1-The Ohio State University, 2-Alferd , 3-St. Vincent, 4-Hahnemann
- z10 MTX Used as a Graft-Versus-Host- Prophylactic: 1-Yes 0-No

Bone Marrow Transplant Example

- \blacksquare The main endpoint is disease-free survival ($t2$ and d3) for the three risk groups, ALL, AML Low Risk, and AML High Risk.
- We are also interested in possibly using the covariates z1–z10 to adjust for other factors. We can do this with stepwise regression or hand examination of the results of adding or removing variables.
- \blacksquare In addition, the time-varying covariates for acute GVHD, chronic GVHD, and platelet recovery may be useful.

Time-Dependent Covariates

- \blacksquare A time-dependent covariate is one that changes value in the course of the study.
- \blacksquare For variables like age that change in a linear manner with time, we can just use the value at the start.
- But it may be plausible that when and if GVHD occurs, the risk of relapse or death increases, and when and if platelet recovery occurs, the risk decreases.
- We form a variable precovery which is $= 0$ before platelet recovery and is $= 1$ after platelet recovery, if it occurs.
- \blacksquare For each subject where platelet recovery occurs, we set up multiple records (lines in the data frame); for example one from $t = 0$ to the time of platelet recovery, and one from that time to relapse, or death, or end of study.
- We do the same for acute GVHD and chronic GVHD.
- \blacksquare For each record, the covariates are constant.

id group t1 t2 d1 d2 d3 ta da tc dc tp dp 1 ALL 2081 2081 0 0 0 67 1 121 1 13 1

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- Let A, C, and P stand for the event occurs for that patient at some time. Each of the eight possible combinations of A or not-A, with C or not-C, with P or not-P occurs in this data set.
- A always occurs before C and P always occurs before C if both occur; this is for medical reasons.
- \blacksquare Thus there are ten kinds of patients in the data set: None, A, C, P, AC, AP, PA, PC, APC, and PAC.
- **There could be as many as** $1 + 3 + (3)(2) + 6 = 16$
- \blacksquare This is why a package to assist with this is helpful

Possible and Actual Event Sequences

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- Different subjects could have 1, 2, 3, or 4 intervals depending on which of acute GVHD, chronic GVHD, and/or platelet recovery occurred.
- \blacksquare The final interval for any subject has status $= 1$ if the subject relapsed or died at the end of that interval, otherwise the status is 0.
- Any earlier intervals have status $= 0$.
- Even though there might be multiple lines in the data frame, there is never more than one event, so no alterations need be made in the estimation procedures or in the interpretation of the output.
- The function tmerge in the survival package eases the process of constructing the new data frame.

KMsurv bmt data

z1 Patient Age In Years

- z2 Donor Age In Years
z3 Patient Sex: 1-Mal
- Patient Sex: 1-Male, 0-Female
- z4 Donor Sex: 1-Male, 0-Female
- z5 Patient CMV Status: 1-CMV Positive, 0-CMV Negative
- z6 Donor CMV Status: 1-CMV Positive, 0-CMV Negative
- z7 Waiting Time to Transplant In Days
- z8 FAB: 1-FAB Grade 4 Or 5 and AML, 0-Otherwise
- z9 Hospital: 1-The Ohio State University, 2-Alferd , 3-St. Vincent, 4-Hahnemann
- z10 MTX Used as a Graft-Versus-Host- Prophylactic: 1-Yes 0-No

Starting with all these covariates, we eliminated sequentially Patient and Donor Sex, Patient and Donor CMV Status, Waiting time, and MTX.

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Fixed Covariates for the bmt Data

```
require(KMsurv)
require(survival)
data(bmt)
nsubj <- dim(bmt)[1]
id \leftarrow 1:nsubibmt1 <- data.frame(id,bmt) #to identify the subject across multiple lines
bmt1$group <- factor(bmt1$group,labels=c("ALL","AML-Low","AML-High"))
bmt1$z9 <- factor(bmt1$z9) #hospital factor
bmt1.sum \leftarrow with(bmt1, Surv(t2, d3))> drop1(coxph(bmt1.surv~group+z1*z2+z8+z9,data=bmt1),test="Chisq")
Single term deletions
```

```
Model:
bmt1.surv \degree group + z1 * z2 + z8 + z9
      Df AIC LRT Pr(>Chi)
<none> 719.58
group 2 721.76 6.1738 0.0456426 * #ALL, AML-High, AML-Low
z8 1 726.43 8.8504 0.0029303 ** #1-FAB Grade 4 Or 5 and AML, 0-Else
z9 3 725.79 12.2066 0.0067079 ** #Hospital
z1:z2 1 729.23 11.6537 0.0006407 *** #Patient Age by Donor Age interaction
                                             \Rightarrow298
```

```
> summary(coxph(bmt1.surv~group+z1*z2+z8+z9,data=bmt1))
Ca11:coxph(formula = bmt1.surv \degree group + z1 * z2 + z8 + z9, data = bmt1)
```

```
n= 137, number of events= 83
```


We will use the two age variables and FAB score in the following. We omit the hospital effect since the significance test is possibly invalid (hospital-level effect, not patient effect).

```
> summary(coxph(bmt1.surv~group,data=bmt1))
                coef exp(coef) se(coef) z Pr(>|z|)<br>5742 0 5632 0 2873 -1 999 0 0457 *
groupAML-Low -0.5742groupAML-High 0.3834 1.4673 0.2674 1.434 0.1516
> summary(coxph(bmt1.surv~group+z8,data=bmt1))
                 \c{o} exp(coef) se(coef) z Pr(>|z|)groupAML-Low -0.90450 0.40475 0.32031 -2.824 0.00475 **
groupAML-High -0.05195 0.94938 0.32060 -0.162 0.87128
z8 0.76950 2.15868 0.27032 2.847 0.00442 **
```
With group alone, AML-High is riskier than ALL and AML-Low is less risky. The FAB variable z8, which is 1 only for AML, 1/3 of the AML-Low cases and 60% of the AML-High cases, this absorbs some of the risk of the riskiest AML cases, so that the group effect shows both AML groups as less risky than ALL.

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> newgroup <- unclass(bmt1\$group)+bmt1\$z8*3 #five different numerical values > with(bmt1,table(unclass(group)+z8*3))

```
1 2 3 5 6
38 36 18 18 27
> with(bmt1,table(group,z8))
         z8
group 0 1
 ALL 38 0
 AML-Low 36 18
 AML-High 18 27
> newgroup <- factor(newgroup,
  labels=c("ALL","AML-Low","AML-High","AML-Low+FAB","AML-High+FAB"))
> summary(coxph(bmt1.surv~newgroup,data=bmt1))
```
 \c{o} exp(\c{o} ef) se(\c{o} ef) z Pr($>|z|$) newgroupAML-Low -0.7759 0.4603 0.3384 -2.293 0.02185 * newgroupAML-High -0.2144 0.8070 0.3791 -0.566 0.57172 newgroupAML-Low+FAB -0.2829 0.7536 0.3653 -0.774 0.43868 newgroupAML-High+FAB 0.7935 2.2112 0.2903 2.734 0.00626 **

```
> AIC(coxph(bmt1.surv~newgroup,data=bmt1))
[1] 731.9691
> AIC(coxph(bmt1.surv~group+z8,data=bmt1))
 Score separately<br>\longleftrightarrow \longleftrightarrow \longleftrightarrow \Rightarrow \Rightarrow \Rightarrow \Diamond \Diamond
```
Construction of TDC Data Set

Using tmerge we set up the time-dependent covariates data set.

bmt2 <- tmerge(bmt1,bmt1,id=id,tstop=t2) #sets up new data set bmt2 <- tmerge(bmt2,bmt1,id=id,agvhd=tdc(ta)) #adds aghvd as tdc
bmt2 <- tmerge(bmt2,bmt1,id=id,cgvhd=tdc(tc)) #adds cghvd as tdc bmt2 <- tmerge(bmt2,bmt1,id=id,cgvhd=tdc(tc)) #adds cghvd as tdc $bmt2 \leftarrow \text{terge}(bmt2, bmt1, id=id, precovery = tdc(tp))$

status <- as.integer(with(bmt2,(tstop==t2 & d3)))

status only = 1 if at end of t2 and not censored

bmt2 <- data.frame(bmt2,status)

bmt2.surv <- with(bmt2,Surv(time=tstart,time2=tstop,event=status,type="counting"))

#counting process formulation of Surv

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Add Time-Dependent Covariates

> summary(coxph(bmt2.surv~group+z1*z2+z8+agvhd+cgvhd+precovery,data=bmt2))

n= 341, number of events= 83

Neither acute GVHD nor chronic GVHD has a statistically significant effect here or in a model with the other one removed. Platelet recovery is highly significant. > summary(coxph(bmt2.surv~group+z1*z2+z8+precovery,data=bmt2))

n= 341, number of events= 83

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Model Checking

We can use all the same tools for model checking in data sets with time dependent covariates as we do with data sets with only fixed covariates. This includes

- **Schoenfeld residuals correlated with "time" to test for** proportionality of hazards.
- **Martingale residuals plotted vs numeric covariates to check for** functional form.
- Martingale residuals and deviance residuals plotted vs the linear predictor to identify possible outliers.
- Columns of dfbeta to identify possible influential points: points whose removal changes the fit importantly.

We won't use the Cox-Snell residuals since this plot has low capacity to detect problems. Ω The original data set is 137 rows and 22 columns, corresponding to 137 patients with a number of events that depends on the type of event:

Model checking when using the original data set is as we have seen before.

Model Checking

 $55/120 = 45.8\%$ Survival rate with precovery $1/17 = 5.9\%$ Survival rate without precovery

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The original data set is 137 rows and 22 columns, corresponding to 137 patients. The data set for time-dependent analysis is 341 rows by 29 columns. This means that there are 341 different patient by time-dependent covariate intervals, about an average of 2.5 intervals per patient. The first extra column is id one unique value per patient, and the others are tstart, tstop, delimiting the intervals, agvhd, cgvhd, precovery, stating which events have already occurred before that interval, and status indicating whether the interval terminates with recurrence or death.

An argument to the residual command is collapse which has the default value collapse = $F = FALSE$ which gives us 341 residuals or collapse = id which combines all the residuals for each patient, resulting in 137 residuals. Both approaches can be useful. The first gives us one residual per patient per values of the time-dependent covariates and the second has one residual per patient. If plotted vs. something in the data set it has to be from bmt2 in the first case and bmt1 in the second, even though the residual vector is derived from the model using the data set bmt2.

Schoenfeld Residuals

```
bmt2.cox <- coxph(bmt2.surv~group+z1*z2+z8+precovery,data=bmt2)
bmt2.zph <- cox.zph(bmt2.cox)
print(bmt2.zph)<br>plot.zph <- function(i,df=4){
                                 #df = 4 is the default degree of the spline
 plot(bmt2.zph[i],df=df) #df = 2 uses linear splines
}
         chisq df p
group 1.0458 2 0.59 #Disease
z1 0.6625 1 0.42 #Patient Age
z2 2.3980 1 0.12 #Donor Age
z8 0.3216 1 0.57 #FAB Score
precovery 0.0721 1 0.79 #Platelet Recovery
z1:z2 0.9210 1 0.34 #Age Interaction
GLOBAL 6.3820 7 0.50 #No major signs of non-proportionality
pdf("Schoenfeld3.pdf") #These are for z2 = donor age
plot.zph(3) #This is column 3/7 of the scaled schoenfeld resids
dev.off()
pdf("Schoenfeld3a.pdf")
plot.zph(3,df=2)
dev.off()
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Martingale Residuals

```
plot.mres.z1 <- function(){
 mres <- residuals(coxph(bmt2.surv~group+z2+z8+precovery,data=bmt2),
     type="martingale")
  plot(bmt2$z1,mres,xlab="Patient Age",ylab="Martingale Residuals")
  lines(lowess(bmt2$z1,mres))
  title("Martingale Residuals vs. Patient Age")
}
plot.mres.z2 <- function(){
 mres <- residuals(coxph(bmt2.surv~group+z1+z8+precovery,data=bmt2),
     type="martingale")
  plot(bmt2$z2,mres,xlab="Donor Age",ylab="Martingale Residuals")
  lines(lowess(bmt2$z2,mres))
  title("Martingale Residuals vs. Donor Age")
}
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Martingale Residuals

```
plot.mres.z12 <- function(){
 mres <- residuals(coxph(bmt2.surv~group+z1+z2+z8+precovery,data=bmt2),
     type="martingale")
 plot(bmt2$z1*bmt2$z2,mres,xlab="Patient Interaction",
    ylab="Martingale Residuals")
 lines(lowess(bmt2$z1*bmt2$z2,mres))
 title("Martingale Residuals vs. Patient Interaction")
}
plot.mres.z7 <- function(){
 mres <- residuals(coxph(bmt2.surv~group+z1*z2+z8+precovery,data=bmt2),
     type="martingale")
 plot(bmt2$z7,mres,xlab="Waiting Time",ylab="Martingale Residuals")
 lines(lowess(bmt2$z7,mres))
 title("Martingale Residuals vs. Waiting Time")
}
```
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Martingale Residuals vs. Patient Age

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Martingale Residuals vs. Donor Age

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Martingale Residuals vs. Patient Interaction

Patient Interaction

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Martingale Residuals vs. Waiting Time

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Martingale and Deviance Residuals

```
bmt2.mart <- residuals(bmt2.cox,type="martingale")
bmt2.dev <- residuals(bmt2.cox,type="deviance")
bmt2.dfb <- residuals(bmt2.cox,type="dfbeta")
bmt2.preds <- predict(bmt2.cox)
plotr.mart <- function(){
 plot(bmt2.preds,bmt2.mart,xlab="Linear Predictor",ylab="Martingale Residual")
 title("Martingale Residuals vs. Linear Predictor")
}
plotr.dev <- function(){
```

```
plot(bmt2.preds,bmt2.dev,xlab="Linear Predictor",ylab="Deviance Residual")
 title("Deviance Residuals vs. Linear Predictor")
}
```
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Three smallest martingale residuals are from patient id's 14, 100, and 103.

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> bmt1[c(14,100,103),imp.vars1] id group t1 t2 d1 d2 d3 ta da tc dc tp dp z1 z2 z8 14 14 ALL 1167 1167 0 0 0 39 1 487 1 1167 0 27 22 0

100 100 AML-High 2024 2024 0 0 0 2024 0 180 1 16 1 35 41 1 103 103 AML-High 845 845 0 0 0 845 0 845 0 20 1 40 39 1

Patient 14 is in the medium-risk group, had a long survival time (censored), but early AGVHD and CGVHD, and no platelet recovery. Patients 100 and 103 are in the highest risk-group, had long survival times (censored), and early platelet recovery.

Deviance Residuals vs. Linear Predictor

No unusualy low or high deviance residuals.

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DFBETA

The residuals = dfbeta matrix is 341 by 7 with rows corresponding with patient \times intervals and columns corresponding to the coefficients groupAML-Low, groupAML-High, z1, z2, z8, precovery, z1:z2.

dfBeta vs. Observation Order

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dfBeta vs. Observation Order

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dfBeta vs. Observation Order

Observations 88 and 128 high and 14, 84, and 116 low.

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dfBeta vs. Observation Order

Observations 84 and 129 high and 116 and 118 low.

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dfBeta vs. Observation Order

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dfBeta vs. Observation Order

Observation 14 high and 30, 36, 77, 85, 86, 87 low.

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dfBeta vs. Observation Order

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Observations 116 and 118 have very young patient/donor combinations. These are extreme in the linear function of age and especially in the product. Observations 128 and 129 are in AML-High but no z8 FAB extra risk and have very early deaths. Observations 84–87 have the lowest risk group, $AML-Low + no$ extra FAB risk, but early deaths. Observation 88 is a low risk (of progression-free survival) with early platelet recovery but relapsed at

a long interval.

This kind of analysis can identify errors. It can identify problems like use of linear age. Some outliers are explicable from unusual predictive values. The plots we use can identify these unusual combinations much more easily than just staring at the data.

This kind of analysis is even more important in early stages of the project because it can identify specious observations as well as influential ones.

Recurrent Events

- Sometimes an appropriate analysis requires consideration of recurrent events.
- \blacksquare A patient with arthritis may have more than one flareup. The same is true of many recurring-remitting diseases.
- \blacksquare In this case, we have more than one line in the dataframe, but each line may have an event.
- We have to use a "robust" variance estimator to account for correlation of time-to-events within a patient.

Bladder Cancer Data Set

The bladder cancer dataset from Kleinbaum and Klein contains recurrent event outcome information for eighty-six cancer patients followed for the recurrence of bladder cancer tumor after transurethral surgical excision (Byar and Green 1980). The exposure of interest is the effect of the drug treatment of thiotepa. Control variables are the initial number and initial size of tumors. The data layout is suitable for a counting processes approach.

This drug is still a possible choice for some patients. Another therapeutic choice is Bacillus Calmette-Guerin (BCG), a live bacterium related to c[ow](#page-48-0) [t](#page-50-0)[u](#page-48-0)[b](#page-49-0)[e](#page-50-0)[rc](#page-0-0)[ul](#page-59-0)[os](#page-0-0)[is.](#page-59-0) Ω

Bladder Cancer Data Set

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- There are 85 patients and 190 lines in the dataframe, meaning that many patients have more than one line.
- **Patient 1 with 0 observation time was removed.**
- Of the 85 patients, 47 had at least one recurrence and 38 had none.
- **18 patients had exactly one recurrence.**
- \blacksquare There were up to 4 recurrences in a patient.
- Of the 190 intervals, 112 terminated with a recurrence and 78 were censored.
- Different intervals for the same patient are correlated.
- Of the 85 patients, 47 had at least one recurrence and 38 had none.
- Of the 190 intervals, 112 terminated with a recurrence and 78 were censored.
- If Is the effective sample size 47 or 112? This might narrow confidence intervals by as much as a factor of $\sqrt{112/47} = 1.54$
- What happens if I have 5 treatment and 5 control values and want to do a t-test and I then duplicate the 10 values as if the sample size was 20? This falsely narrows confidence intervals by a factor of √ $'2 = 1.41$. Ω

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```
require(survival)
vars <- c("id","status","interval","intime","tstart","tstop","tx","num","size")
bladder <- read.table("bladder.dat",header=F,col.names=vars)
bladder <- bladder[-1,] #remove subject with 0 observation time
```
#bladder.dat from Kleinbaum and Klein with lines before and after data removed

bladder.surv <- with(bladder,Surv(time=tstart,time2=tstop,event=status, type="counting"))

bladder.cox1 <- coxph(bladder.surv~tx+num+size,data=bladder) #biased variance co-variance matrix

bladder.cox2 <- coxph(bladder.surv~tx+num+size+cluster(id),data=bladder) #unbiased though this reduces power

bladder.cox3 <- coxph(bladder.surv~tx+num+cluster(id),data=bladder) #remove non-significant size variable

```
> summary(bladder.cox1)
Call:
cosph(formula = bladder.surv * tx + num + size, data = bladder)n= 190, number of events= 112
        \c{o} exp(coef) se(coef) z Pr(>|z|)tx = 0.41164 0.66256 0.19989 -2.059 0.039466 *
num  0.16367  1.17782  0.04777  3.426  0.000611 ***
size -0.04108  0.95975  0.07029 -0.584  0.558967
> summary(bladder.cox2)
Ca11:cosh(formula = bladder.surv * tx + num + size + cluster(id),data = bladder)n= 190, number of events= 112
        coef exp(coef) se(coef) robust se z Pr(>|z|)
tx -0.41164 0.66256 0.19989 0.24876 -1.655 0.09798 .
num  0.16367  1.17782  0.04777  0.05842  2.801  0.00509   **
size -0.04108 0.95975 0.07029 0.07421 -0.554 0.57991
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```
> summary(bladder.cox1)
```

```
Concordance= 0.624 (se = 0.03)
Rsquare= 0.074 (max possible= 0.992 )
Likelihood ratio test= 14.66 on 3 df, p=0.002127Wald test = 15.9 on 3 df, p=0.001187Score (logrank) test = 16.18 on 3 df, p=0.001042> summary(bladder.cox2)
Concordance= 0.624 (se = 0.03)
Rsquare= 0.074 (max possible= 0.992 )
Likelihood ratio test= 14.66 on 3 df, p=0.002127Wald test = 11.19 on 3 df, p=0.01073Score (logrank) test = 16.18 on 3 df, p=0.001042, Robust = 10.84 p=0.01263
```
(Note: the likelihood ratio and score tests assume independence of observations within a cluster, the Wald and robust score tests do not).

```
> round(bladder.cox2$naive.var,4)
       [0.1] [0.2] [0.3][1,] 0.0400 -0.0014 0.0000
[2,] -0.0014 0.0023 0.0007
[3,] 0.0000 0.0007 0.0049
> round(bladder.cox2$var,4)
       [0,1] [0,2] [0,3][1, 1 \ 0.0619 - 0.0026 - 0.0004][2,] -0.0026 0.0034 0.0013
[3,] -0.0004 0.0013 0.0055
> sqrt(with(bladder.cox2,diag(var)/diag(naive.var)))
[1] 1.244492 1.223092 1.055761
```
These are the ratios of correct confidence intervals to naive ones.

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```
> summary(bladder.cox3)
Ca11:coxph(formula = bladder.surv \tilde{t} tx + num + cluster(id), data = bladder)
 n= 190, number of events= 112
       \c{coeff}\, \exp(\, \c{coeff})\, \sec(\, \c{coeff})\, \text{robust}\, \sec \, z\, \Pr(\, > |z|)\,tx -0.41172 0.66251 0.20029 0.25153 -1.637 0.10166
num 0.17001 1.18531 0.04646 0.05636 3.016 0.00256 **
---
Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
   exp(coef) exp(-coef) lower .95 upper .95
tx 0.6625 1.5094 0.4047 1.085
num 1.1853  0.8437  1.0613  1.324
Concordance= 0.623 (se = 0.029 )
Rsquare= 0.073 (max possible= 0.992 )
Likelihood ratio test= 14.31 on 2 df, p=0.0007799
Wald test = 10.24 on 2 df, p=0.005969Score (logrank) test = 15.81 on 2 df, p=0.0003696, Robust = 10.6 p=0.005001
  (Note: the likelihood ratio and score tests assume independence of
    observations within a cluster, the Wald and robust score tests do not).
```
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